

Legume-supported cropping systems for Europe

Legume Futures Report 1.3

Novel feed and non-food uses of legumes

Compiled by:

F.L. Stoddard

University of Helsinki

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Legume Futures

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FOREWORD

The legume family, *Fabaceae*, is one of the largest in the plant kingdom. Almost all species in the family form symbioses with bacteria in the family *Rhizobiaceae*, leading to biological nitrogen fixation. While some other species scattered through the plant kingdom also fix nitrogen through symbioses, the legumes are the largest group and the most useful in agriculture.

This capacity for nitrogen fixation has several impacts. It means that the plants can grow in nitrogen deficient soils and at the same time produce protein-rich plant material, particularly protein rich seeds. This high protein content and production, which is intrinsic to legumes, determines much of the role of legumes not only in general human and animal nutrition, but also their suitability for novel feed uses and uses in the non-food sector. Biological nitrogen fixation is a characteristic of pioneer plants and so gives rise to another potential use of legumes in the bioremediation or colonization of soils otherwise unsuited for agriculture.

Legumes are also, compared with cereals, rich in a range of secondary plant compounds. Legumes have evolved mechanisms to produce and concentrate these compounds to protect against pest and disease attack. The bioactivity of these compounds opens up non-food opportunities which are specific to legumes.

This report also looks at non-traditional feed uses, such as whole-crop silage and fish feeds, examines some industrial uses of legumes in the bio-based economy, and concludes with a catalogue of recent demonstrations of the activities of bioactive compounds derived from legumes. A comprehensive gathering of such data would require hundreds of pages and thousands of references, and this document is intended to introduce the reader to the literature and present some of the more interesting highlights that are relevant in the context of European agriculture.

Fred Stoddard,

Helsinki, Finland, October 2013

INTRODUCTION

This report considers novel uses of legumes in the animal feed and non-food industries. If we are to consider "novel" uses, first we need to define "traditional" uses. With regard to animal feed, traditional uses include the grazing of pastures or saving of hay from grass mixed with legumes including clovers and alfalfa. Other traditional uses include usage of soya bean meal as a protein supplement for poultry, pigs and ruminants, and most uses of dried pea, faba bean or lupins for the same purpose. Lupins, being a fairly recently developed set of crops, may be considered novel in some regions and markets. Less traditional uses of legumes include using whole-crop grain legumes for silage, feeding lupin seeds to non-ruminants, and any use of plant proteins for feeding fish.

Among non-food uses, green manuring is too familiar to need mention here. The capacity of legume-rhizobium symbioses to fix atmospheric dinitrogen leads to potential impacts in bioenergy and biorefining that are discussed below. Legumes contain many secondary compounds that both contain nitrogen and protect the plant from herbivores seeking nutrition. Some of these compounds have been used in traditional medicine, and modern science has validated some of these properties. Other medicinal, biocidal or growth-promoting uses have been discovered.

There is a huge and developing literature on the non-traditional usage of legumes in foods, such as the substitution of other legume "milks" for soya milk, itself a substitution for milk from cows. While this literature is not addressed directly here, we touch on peptides and some other bioactive compounds that would most likely be consumed in a food matrix.

Three reviews have provided good examples and starting points for this review. Morris (1997) reviewed industrial and pharmaceutical legumes, and updated the review in 2003, with a preference for species adapted to growing in the United States. Howieson et al. (2008) reviewed a selected range of uses of legumes, focusing on human-health benefits, anthelminthics for ruminants, aquaculture, and deep rooting for access to water and nutrients.

NOVEL USES IN ANIMAL FEED

F.L. Stoddard, Department of Agricultural Sciences, University of Helsinki, Finland

There is a small but positive literature on ensiling of grain legumes and on their use as protein supplements for ruminants. Legumes have a low content of water-soluble carbohydrates and a high buffering capacity, so require wilting or treatment with additives such as formic acid or lactic acid bacteria in order to ensile satisfactorily (Pursiainen & Tuori 2008). Given these restrictions, silage can be made from whole-crop pea (Pursiainen & Tuori 2008, Borreani et al. 2009), faba bean (Pursiainen & Tuori 2008, Borreani et al. 2009, Pakarinen et al. 2011), white lupin (Fraser et al. 2005a, Pakarinen et al. 2011), narrow-leafed lupin (Fraser et al. 2005b) and yellow lupin (Serrano 1989). Results are often improved by mixing a cereal with the legume, e.g., durum wheat - faba bean (Mariotti et al. 2012). Soya bean meal as a protein supplement can be replaced in dairy or beef cattle feeding by seed or meal of pea (Brunschwig & Lamy 2003), faba bean (Moss et al. 2000), white lupin (Froidmont & Bartiaux-Thill 2004), narrow-leafed lupin (Eriksson 2010, Niwinska & Andrzejewski 2011) and yellow lupin (Marley et al. 2008). The choice of legume for the supplement depends on the background silage, as the slow breakdown of lupins makes them more suitable with grass silage, while pea and faba bean are more suitable with maize silage (Wilkins & Jones 2000).

The presence of anti-nutritional factors in grain legumes has more effect on their use for monogastrics than on that for ruminants. Trypsin inhibitors, vicine-convicine, tannins and alkaloids are implicated as limiting factors, depending on the animal and the feed. Nevertheless, reports can be found on the successful use of almost every grain legume species for feeding pigs, broiler hens, laying hens, and turkeys. Laying hens are notoriously sensitive to the vicine-convicine of faba bean, and broilers also benefit from the absence of these anti-s (Vilarino et al. 2009). Normal-vicine faba bean was acceptable at up to 31% in a mixed feed for broilers (Laudadio et al. 2011) and white lupin was acceptable up to 24% (Laudadio & Tufarelli 2011). Field pea, in contrast, could be included up to 50% of the feed of laying hens (Fru-Nji et al. 2007). The low digestibility of the storage galactan in lupin seeds reduces their value in monogastric feeds. Supplementing a feed based on yellow lupin (Olkowski et al. 2010) or narrow-leafed lupin (Steenfeldt et al. 2003) with a glycanase mixture resulted in significantly improved performance of broilers. Pea was more digestible than faba bean and narrow-leafed lupin in turkey diets (Palander et al. 2006). Pea and faba bean are equally suitable for grower and finisher pigs (Smith et al. 2013), and low tannin content provides a valuable increase in the apparent metabolizable energy of the legume (Crépon et al. 2010). The standard ileal digestibility of protein from narrow-leafed and yellow lupin was as good as that of soy bean meal, while those of pea and faba bean were significantly lower (Jezierny et al. 2011).

Thus European cool-season grain legumes can be used to replace part or all of the imported soya bean meal used in feeding poultry, pigs and ruminants, at least on an experimental scale. Amino acid balance, ileal digestibility for monogastrics, rumen stability for ruminants, apparent metabolisable energy, and the restrictive effects of the individual antinutritional factors of each cultivar, all need to be taken into account. Some of these limiting factors can be amended through breeding, others through post-harvest treatment, and others by using additives or other means of balancing the rest of the feed formulation.

LEGUMES IN FEEDS FOR FISH AND CRUSTACEANS

F.L. Stoddard, Department of Agricultural Sciences, University of Helsinki, Finland;P.P.M. Iannetta, A.J. Karley, and G. Ramsay, The James Hutton Institute, Dundee, UK;Z. Jiang, Department of Food and Environmental Sciences, University of Helsinki, Finland;J.G. Bell, and D.R. Tocher, Institute of Aquaculture, University of Stirling, UK; andV. Crampton, EWOS Innovation, Sandnes, Norway

Aquacultural production is an important economic activity in the EU, with a total value estimated at €2,800 million, representing 7.6% of global aquaculture value and 4.6 % of global tonnage. This represents almost a doubling in output since the late 1980s (Tacon, 1997). The enterprises driving this production vary in scale from cottage industry to large multinational operations, although 90 % are small-to-medium enterprises (SMEs) (Varadi et al., 2000).

The farming of fish and shellfish for food supplies approximately 50 % (by mass) of all the fish and shellfish consumed worldwide (National Research Council, 2011), and the bulk of this production originates from the highly commercialised sector that has transformed the practice of aquaculture largely through the use of modern science and engineering. Commercial aquaculture production is a high-value activity and wholly dependent on the use of manufactured feed. The sourcing of the components of this feed is of primary consideration to achieve efficient production while minimising environmental impacts. The increased output of the commercial aquaculture sector with reduced environmental impacts has been achieved by reducing the use of wild fish resources in feed and increasing feed conversion efficiency. In particular, the amount of high-quality edible fish produced per unit of fish used in fish-feed has increased.

Aquaculture also provides rural employment and alleviates rural poverty, and sustainability of production is now considered a more desirable objective than volume of production (De Silva 2001, Foresight 2011). Moreover, as the world's population is projected to be 9 billion by 2050 (United Nations, 2009), there is doubt in equal measures about the ability of both traditional cottage and commercial aquaculture systems to meet increasing global demand for food. Although the aquaculture industry produces a vast array of fin fish, molluscs and crustaceans by catch and managed activities, two groups of farmed species, namely shrimp or prawns (Penaeidae) and the salmon family (Salmonidae), offer opportunities for increased economic potential and enhanced food security, due to their relatively high market value and potential for vertical business integration (Swick & Cremer, 2001). Most importantly, although they are omnivorous to carnivorous, research has shown that other sources of protein and oil can substitute for the fish products normally (and in natural ecosystems) used for their nutrition (Trushinski et al., 2006). Vegetarian fish, such as carp, may be able to make even better use of plant-based feeds. Α successful future for European aquaculture industries appears to rest on an abundant, affordable and, most importantly, sustainable supply of alternative to fish meal (FM) in

aquaculture feeds. The ingredients that can substitute for fish meal may be sourced from land animals, microorganisms (algae, yeast and bacteria) and plant sources such as oilseeds, cereals and grain legumes (Gatlin et al., 2007). The feed is generally formulated into pellets that are sprinkled onto the surface of the water, from which the fish rapidly snatch it. Cohesion of the pellets is essential, so components such as starch that gelatinizes in the pellet extruder, and wheat gluten are commonly used (Brinker & Friedrich, 2012).

Increasing the amount of fish produced per unit of fish used in feed is central to developments in the nutrition of fish. This requires increasing the plant-based components of fish feed while not compromising fish growth, health and product quality. However, there have been few empirical determinations of the nutrient requirements of fish species farmed using plant-based feeds (National Research Council, 2011). Plant feedstuffs for aquaculture tend to provide relatively low levels of essential amino acids, particularly methionine, tryptophan and histidine, but these can be supplemented by inexpensive, synthetic amino acids (Dias et al., 2005). Higher plants also provide low levels of the bioactive long-chain, polyunsaturated (omega-3) fatty acids that are produced by marine algae. Hence, fish meal and fish oil are still important components of many aquaculture feeds, but research on plant oils is moving rapidly and plant sources of long-chain, polyunsaturated fatty acids are being developed to deliver quantities at the commercial scale necessary to meet the demands of the aquaculture industry (Miller et al., 2008; Naylor et al., 2009).

Initial success using faba bean and narrow-leafed lupin as aquaculture feedstocks, and the increased global demand for fish produced in Europe, may help to develop the market for locally produced grain legumes in Europe. Some large commercial fish farms (such as EWOS, UK) have committed to avoid purchasing soya bean grown in previously forested areas of the tropics. In experiments, soya bean meal had a negative and dose-dependent effect on fish digestive systems attributed to anti-nutritional factors (ANFs), with a consequent loss of fish health, meat quality and yield (Krogdahl et al., 2003). Thus heat treatments to denature the ANFs are usually necessary when soya bean meal is to be used for fish feed, and the economic and environmental costs of fish meal, soya bean meal, and alternative grain legume protein sources need to be taken into the overall equation.

As shown below, faba bean and narrow-leafed lupin are both particularly well suited for use in salmon feed, and dehulled, protein enriched meal could successfully displace other vegetable protein sources, such as those based on soya bean. Formulation of fish feed is as complex and precise as that of other animal feeds, and in experiments, feeds under comparison are formulated to be as isoenergetic and isonitrogenous as possible.

Grain legumes contain anti-nutritional factors that affect different animal systems, including proanthocyanidins and condensed tannins, along with alpha-galactosides, lectins, saponins, phytic acid, and trypsin inhibitors. In faba bean, there are also the pyrimidine glycosides vicine and convicine and the free amino acid L-DOPA (L-3,4-dihydroxyphenylalanine), while lupins contain alkaloids.

The proanthocyanidins and condensed tannins that reduce the efficiency of the digestion of proteins by animals (Griffiths 1986; Griffiths and Jones 2006) are in the seed coat, and hence removed in the dehulling and air-classification processes that are expected to be used in making protein concentrates for fish feed (see below). Most cultivars of narrowleafed lupin are low in tannin, and two zero-tannin genes (zt1 and zt2) are used in faba bean breeding programmes (Crépon et al., 2010). Of the remaining anti-nutritional factors in faba bean, vicine and convicine are associated with protein bodies and are likely to have elevated concentration in purified protein extracts (Olsen & Andersen, 1978). Their aglycones, divicine and isouramil, cause oxidative stress in rats, laying hens, and glucose-6-phosphate dehydrogenase deficient humans (Arese et al. 1981), and are considered the most likely of the faba bean anti-nutritional factors to affect fish. A gene for low vicineconvicine content, zvc, has been identified and used in faba bean breeding programmes (Crépon et al., 2010), so it is likely that this factor will cease to be a significant problem in the near future. ANFs are among the natural defence mechanisms of plants, and they are varied in chemistry and in mode of action. Their content can often be reduced by breeding in the long term and by processing in the short term, although the processing is generally expensive and complete removal is seldom commercially feasible. More significant is the fact that salmonid growth and feed efficiency tends to be higher on moderate levels of several plant proteins than on a single, perhaps because in this way no single ANF is limiting. This observation is of critical importance for feed formulations, which demand that that many new plant protein sources have a role alongside those already used in the blend. A complex blend of several legume components replaced 66% of the dietary animal protein without significantly reducing growth or quality of the rainbow trout (Gomes et al. 1995).

There are several times more publications on the use of lupins than that of faba bean in feeds for salmonids. A feed based on soya bean (plant protein replacing 20% of animal protein) was significantly worse for Atlantic salmon, *Salmo salar*, than other nine feeds, and it was associated with symptoms of enteritis, whereas feeds based on pea or dehulled faba bean gave the best results (Aslaksen et al. 2007). The low cellulose content of the dehulled narrow-leafed lupin was associated with high lipid digestibility (Aslaksen et al. 2007). Narrow-leafed lupin kernel meal showed no negative effects on the growth of rainbow trout at the maximum experimental level of 30% of the diet (Glencross et al. 2008). Protein concentrate from narrow-leafed lupin resulted in a higher feed conversion ratio than the equivalent from pea (Zhang et al. 2012). Removal of the oligosaccharides

is a necessary step in preparing digestible feeds from narrow-leafed lupin (Glencross et al. 2003).

Replacing animal protein with plant protein results in friability of the fish faeces, and addition of guar gum, extracted from another legume, the guar bean (*Cyamopsis tetragonoloba*), to the feed of rainbow trout allowed the faeces to retain their stability and sink to the bottom of the tank, maintaining system hygiene (Brinker & Friedrich 2012).

Given that sustainably produced plant-derived feedstuffs are available, their suitability to underpin the production of fish species beyond salmonids will also be important, since commercially successful farms for non-salmonids such as turbot, eel, European sea-bass and gilthead sea-bream exist in Europe. There is also considerable additional commercial opportunity to develop the use of grain legume protein in feeds for a wide variety of farmed marine and freshwater species. It is also pertinent to note that carp raised in the earthen ponds that dominate the aquaculture production of Eastern and Central Europe (5% of EU aquaculture output: Varadi et al., 2000) are herbivorous or omnivorous species and easily assimilate protein from grain legumes, including pea, soya bean (Davies & Gouveia 2010), white lupin, yellow lupin and faba bean (Mazurkiewicz 2009), with an optimum protein composition around 30% plant protein and 70% animal protein. European sea bass (Dicentrarchus labrax) showed little difficulty with diets where almost all fish meal was replaced with a mixed plant-based diet containing maize gluten, wheat gluten, soya bean meal and rapeseed meal (Kaushik et al. 2004).

Lupin kernel meal has been widely tested as a feed component for the black tiger shrimp, *Penaeus monodon*, and the Pacific white shrimp, *Litopenaeus vannamei*. In most studies, the kernels have been dehulled and defatted before use. Alkaloids at normal levels caused a brief interruption in feeding by black tiger shrimp, without long-term detriment (Smith et al. 2007a). Depending on the lupin and the shrimp species, the shrimp grew well with up to 40% (Smith et al. 2007b), 41.5% (Smith et al. 2007a), or 50% (Molina-Poveda 2013) of the protein being plant-derived.

The high concentration of protein in aquaculture feeds (with a weighted average of ca. 35 % for salmon), cannot be achieved by most legume grains without further processing. While the main energy component of legume grains is lipid, non-starch polysaccharide (in lupins), or starch, for those legume grains containing starch (such as faba beans), air classification (AC; also known as "air fractionation" or "protein enrichment") may be applied (Shapiro and Galperin, 2005). AC allows the dry separation of particles from finely ground flours according to their size, shape, density and, hence, flow behaviour in an air-stream (Boye et al. 2010; Pelgrom et al. 2013). The fine grinding can be achieved by well controlled pin milling and jet milling methods. After the fine grinding, the protein bodies of the seeds can be detached from the starch granules. Then the protein-rich fraction

containing mainly fine particles that fly faster is separated from the starch-rich fraction containing mainly coarse particles by a stream of air.

As compared to the more traditional protein-starch separation method, wet fractionation, AC is more cost effective (saving energy, water and time), hygienic and ecological. Moreover, AC keeps the processed starch and protein in a more native form without reducing their functionality. The wet-process method is dependent on the solubility difference between protein and starch. A large amount of water is used in wet processing for starch production from pulses such as mung bean and faba bean. The process takes a long time and uses considerable energy input during the washing, delivering and precipitating (centrifuging) steps. In addition, the collection of protein from the wet process needs acid for protein precipitation and further inputs of energy for centrifugation and drying. The heat-drying step in wet-processing of legume protein may denature the proteins and reduce their rehydration capability. As a result, in practice, it is considered difficult to use the protein-containing "wastewater" from wet-processing of legume starch for feed and food (Ishida et al. 2012).

AC has been used for many food and feed purposes in cereal and legume processing, for example, starch purification (Vasanthan & Bhatty 1995, Létang et al. 2001), protein concentration (Gunawardena et al. 2010) and fibre enrichment (Ferrari et al. 2009). The efficiency of AC depends on the nature of the grains and some parameters of the process. First, lipid removal is normally favourable for the protein, fibre and starch separation in lipid-containing materials such as oats (Sibakov et al. 2010) and rapeseed meal (King & Dietz 1987), and hence probably for separating protein from fibre in lupins, but is unnecessary for legumes like faba bean that contain little lipid. Second, lower seed moisture content results in a higher protein-starch separation efficiency of faba bean and field pea (Tyler & Panchuk 1982). Third, the milling should not be too coarse, too fine or break the starch granules in order to achieve good protein-starch separation (Boye et al. 2010, Pelgrom et al. 2013). In addition, the parameter settings of the air-classifier such as wheel speed and airflow also have to be optimised for an acceptable separation of protein from starch. Separation may be improved by re-milling and reclassification of the coarse (starch-rich) fraction, after which the resultant second fine (protein-rich) fraction is combined with the first protein fraction. Multiple passes give increased protein concentration in the fine fraction but with decreasing benefits with each pass (Wu & Nichols, 2005). The efficiency of AC should be comprehensively evaluated, as a process producing a higher yield of a fraction may result in a lower purity (for example, protein concentration in protein fraction) (Pelgrom et al., 2013). Moreover, on the same raw material, higher efficiency of protein separation (the percentage of the flour protein recovered in the protein fraction) may be associated with lower efficiency of starch separation (Tyler & Panchuk, 1982). Protein separation efficiency by AC was up to 68.0% in pea and up to 69.7% in faba bean (Tyler & Panchuk 1982). Since then, technology has advanced, and protein separation efficiency was up to 76.8 % in a pea AC study (Pelgrom

et al. 2013). The presence of some starch is beneficial to the formation of the feed pellets under heat extrusion, and faba bean starch expands well on gelatinisation, so wheat or other cereal starch can be removed from the formulation.

It is clear that intensive European aquaculture will benefit from a reduced reliance on soy bean and marine-based protein sources for environmental, nutritional and supply reasons. The use of de-hulled faba beans and lupins can provide a protein source that meets this need in a sustainable manner. Air classification is an appropriate method for enhancing the protein content of the faba bean component of the feed mixture.

NON-FOOD USES OF LEGUMES

F.L. Stoddard, Department of Agricultural Sciences, University of Helsinki, Finland

One of the main reasons for using bioenergy is the reduction of greenhouse-gas emissions. The manufacture of synthetic nitrogen fertilizer relies heavily on fossil fuel as a source of energy input, and the use of synthetic and organic nitrogen fertilizer leads to the release of nitrous oxide, a potent greenhouse gas. Hence, the use of nitrogen fertilizer in bioenergy cropping is generally minimized, and legumes have a potential role (Stoddard 2008).

First-generation biofuels

The first-generation biofuels are made using simple technologies in order to replace fossil fuels. Starch from cereals and sugars from sugar crops are fermented to bioethanol to replace or substitute for petrol (gasoline). Seed oil can be converted to biodiesel by transesterification, in which the bond of the fatty acid with a glycerol residue is replaced by one with a methanol residue. Biomethane, produced by digestion of a wide range of organic substraits, is sometimes put into this category, but its production is based more on biomass, so it is handled in the next section of this chapter.

Legume starch can be converted to bioethanol in the same way as cereal starch, but since starchy legumes generally yield much less than cereals and their starch content is lower, it is highly unlikely that this will ever be economic or sustainable.

An early life-cycle analysis of bioenergy production showed that the nitrogen-fixation capacity of soya bean gave it a significant advantage over other oilseeds (Hill et al. 2006). Nevertheless, the use of soya oil for bioenergy is as questionable as the use of any other food or feed material, and other oils have been sought. For the semi-arid zone, two woody species contend: Jatropha curcas L. (Euphorbiaceae) and the Indian native Pongamia pinnata (L.) Pierre (sometimes put in genus Millettia) (Fabaceae). The toxicity of Jatropha (it is known in English as vomit nut) has inspired remarkably heated debate about its appropriateness as a crop for poor farmers, and *Pongamia* grows in similar environments, fixes its own nitrogen, and yields more oil per hectare without toxicity problems (Scott et al. 2008. Biswas et al. 2011). Karanjin, a furanoflavonoid reputed to have pesticidal properties, can be extracted from the oil as a value-added compound (Vismaya et al. 2010). The oil-free meal contains anti-nutritional factors that impede its use as an animal feed (Vinay & Sindhu Kanya 2008), but it can be put through a methane digester to recover further bioenergy, and the nitrogen- and mineral-rich residue used as a fertilizer.

Biomass production

The current focus on biomass is largely for energy purposes. Whole crop digestable biomass can be digested to methane, in a basically low-technology process, replacing natural gas. The lignocellulosic (woody) fraction can be combusted for heat and electricity, or converted to cellulosic ethanol for transport fuel. A further option is pyrolysis to synthesis gas (that can be polymerized over catalysts to form liquid fuels) and biochar (a fine charcoal that is used either as a solid fuel or a non-biodegradable, carbon-sequestering soil amendment).

Trees

The wood of *Acacia* species and the North American native *Robinia pseudoacacia* L. (black locust) has long been used as fuel, and ways to convert it into modern biofuels are under investigation. Life-cycle analysis showed that *Robinia* in the Po valley of Italy was superior to *Eucalyptus globulus* and hybrid poplar in Galicia, Spain, for the production of cellulosic bioethanol, largely because of its nitrogen autonomy (Gonzalez-Garcia et al. 2012). Short-rotation coppicing enhances harvestable yield of *Robinia* (Grunewald et al. 2009), and there is already a large literature on managing the crop in this way.

Black locust survives temperatures as low as -40°C in Canada, so it has widespread potential in Europe. Its main drawback is the production of suckers, giving it potential as an invasive alien. At lower latitudes, in Greece, *Acacia cyanophylla* produced more biomass at lower cost per tonne than *Eucalyptus camaldulensis*, *Populus nigra*, and *Arundo donax* (Tzanakakis et al. 2012). The wood of many acacias, as well as *Robinia*, is dense, very durable, and used for such purposes as fence posts, while that of the Australian native *A. melanoxylon* is highly prized for cabinet-making.

Grass-legume intercrops

Many perennial grasses have shown potential as bioenergy crops, but require some nitrogen fertilization, so the scope for using a legume intercrop has been investigated. Results are often disappointing, for a number of reasons. Crop mixtures are quite difficult to design, as they require compatibility and complementarity in resource acquisition both above and below ground, and in phenology and growth cycles. Furthermore, perennial rhizomatous grasses such as miscanthus (*Miscanthus x giganteus*) that are harvested as dry lignocellulose in the spring conserve much of their nitrogen in the soil-plant system, with the result that fertiliser nitrogen requirements are low.

In middle latitudes of North America, yields of switchgrass (*Panicum virgatum*) were not significantly affected by selected legume intercrops, but nitrogen fertilisation was greatly reduced or eliminated (Wang et al. 2010). In lower latitudes of North America, intercropping of alfalfa (*Medicago sativa* L.) with the switchgrass improved overall yield to that of a highly fertilized grass control at a low-fertility site , but did not affect economic

yield at a high-fertility site (Butler et al. 2013). At high latitudes in Europe, reed canary grass (*Phalaris arundinacea*) requires annual N fertilization, and we have preliminary evidence that this can be met by mixed cropping with *Galega orientalis* (Legume Futures project, in progress). Thus there is clear potential to replace nitrogen fertilization of some energy grasses by a nitrogen-fixing legume intercrop, and this contributes positively to the greenhouse gas mitigation effect of the system.

Biorefining

Biorefining offers a way for combining feed and bioenergy production. Alfalfa leaves or leaf protein can be used for livestock feed and the lignified stems as biofuel (González-García et al. 2010, Kamm et al. 2010). Similar suggestions have been proposed for clover-grass or clover-cereal mixtures (Thomsen & Hauggaard-Nielsen 2008). In such systems, the technical product can be polylactate for biodegradable plastics, instead of bioethanol or similar biofuel. The area was recently and comprehensively reviewed (Jensen et al. 2012). A narrowly defined study based on energy and exergy relationships suggested that legumes have no merit in energy crop production, owing to their lower yields, but its authors acknowledged that environmental impacts were not considered, and the only legumes considered among the 12 bioenergy crops were soya bean and alfalfa (Brehmer et al. 2008). The dry matter yields of alfalfa used in their calculations were remarkably low, at 4.5 t/ha. Thus, the Brehmer et al. study is useful insofar as it raises questions that should be addressed, but its conclusions are based on an imperfect data set.

Other industrial uses of woody legumes include fibre and pulp. Sunnhemp (*Crotalaria juncea*) is so-called because of its technological similarity to hemp. It produces long fibres that can be used in similar ways to hemp or jute (Ingle & Doke 2006). Leucaena (*Leucaena leucocephala*) is often used to provide forage in subtropical areas, but its stems also produce good quality fibre for pulping (Díaz et al. 2007).

Tree legumes that are grown for one purpose may be further exploited for other uses. *Acacia senegal* is grown primarily for the production of gum arabic, but it produces an annual crop of seeds with an oil concentration of around 10% (Nehdi et al. 2012). The oil has a high content of oleic and linoleic acids, making it suitable for some food and industrial purposes when the relatively high content of free fatty acids is removed (Nehdi et al. 2012), and the seeds have a protein content up to 39% (Balogun & Fetuga 1986), although the prevalence of antinutritional factors precludes their use as feed. Seed oils of another source of gum arabic, *Acacia arabica,* contain industrially important cyclopropene and epoxy fatty acids (Hosamani et al. 2002). Similarly, black locust trees produce an abundance of seeds that have potential in the biorefining chain or at least as a feedstock for the methane digester.

Phytoremediation

Phytoremediation is bioremediation with plants, that is, the use of plants and their associated microorganisms to amend polluted or contaminated environments. Other physical or chemical methods of remediation are based on moving the contamination to another location or replacing one difficult chemical with another. Phytoremediation, in contrast, is an environmentally friendly technology, requires minimal resources, preserves natural soil properties, acquires its energy (mainly) from sunlight, achieves high levels of microbial biomass and is low in cost, but is usually slow. Use of contaminated ground for production of food or feed is usually undesirable, but there are few concerns about its use for bioenergy or other industrial purposes. By using polluted soils, which simultaneously undergo bioremediation, for bioenergy or industrial cropping, considerable environmentally excellent results of importance both for nature and society can be achieved.

There is a large literature on the application of legumes to bioremediation of contamination by heavy metals and petroleum products. Bioremediation of oil contamination is based on the enhancement of living conditions for soil microorganisms by exudates and breakdown products of plant roots. The oil raises the C:N ratio of the soil, so legumes are more likely to be successful as they require little soil nitrogen. Polycyclic aromatic hydrocarbons (PAHs) are highly toxic, low mobility, durable residues of oil pollution so their degradation is a high priority. Numerous studies have shown that the presence of plants results in faster degradation of oil residues than in bare soil. Plant cover also reduces erosion and legumes improve soil fertility.

Many papers report the detrimental effects of oil pollution, including PAHs, on germination, shoot dry weight and root dry weight of specific plant species. Only a few of these go further and examine the detailed effects on other important root traits such as root length and surface area. Useful variations in these aspects of root structure were shown in 13 grasses and 8 legumes grown in nitrogen-amended, oil-contaminated soil (Kirkpatrick et al., 2006).

The plant has little direct effect on the degradation of petroleum products. Rather, it is the soil-borne microbes that do the degradation, and their community composition is affected by the plant and its associated rhizosphere bacteria, including rhizobia. In a pot experiment, alfalfa was marginally better than oilseed rape or perennial ryegrass at promoting the degradation of pyrene, a PAH (D'Ovidio et al. 2013). None of the three species took up detectable quantities of pyrene. The combination of the perennial legume *Galega orientalis* and its nitrogen-fixing rhizobia *Rhizobium galegae* has shown potential for oil bioremediation purposes (Suominen et al., 2000; Lindström et al., 2003). The combination is tolerant of oil and in pot experiments, promoted the degradation of motor oil (Jussila et al., 2006; Kaksonen et al., 2006). Field experiments to test the effect of *Galega* on oil-contaminated ground are in progress in the Legume Futures project. Faba bean

has been shown effective in promoting the degradation of crude oil in Kuwaiti sands (Radwan et al., 2000).

The presence of oil contamination apparently enhances the release of root exudates from the plants that in turn enhance the growth of certain soil bacteria. In a split-root experiment, where half of the soil was PAH-contaminated and the other half not, the composition of the soil microbial community was altered in the uncontaminated soil, but only in the legume treatments, not in the grasses (Kawasaki et al. 2012). In contrast, in a Chinese experiment, the grass tall fescue promoted PAH degradation more than alfalfa, but the intercrop was more efficient than either sole crop (Sun et al. 2011). The legume does not need to be alive to affect PAH degradation. Pea straw was much more effective than wheat straw at promoting PAH degradation, as it promoted the growth of a wide range of soil microbes in a pot experiment (Shahsavari et al. 2013).

Heavy metal and metalloid contamination can be bioremediated in several ways. The plant may accumulate the contaminant, allowing it to be taken away and safely disposed (phytoextraction or hyperaccumulation); it may be physiologically tolerant of the contaminant and grow regardless of its presence; or it may exclude the contaminant, so the crop product may be safely used. Again, the role of the soil microbiological community is critical, but there is very little literature to indicate any particular effect of legumes on it, in contrast to the oil-contamination literature. Inoculation of yellow lupin with a heavy metal-tolerant strain of Serratia rhizosphere bacteria reduced translocation of the heavy metals to the shoot and improved overall plant growth (El-Aafi et al. 2012). Tolerance to heavy metal contamination is necessary in rhizobia used in such conditions, and as might be expected, can be found when it is sought (Nonnoi et al. 2012).

BIOACTIVE COMPOUNDS FROM LEGUMES

F.L. Stoddard, Department of Agricultural Sciences, University of Helsinki, FinlandP. de Lajudie, Laboratoire des Symbioses Tropicales et Méditerranéennes, Institut de Recherche pour le Développement, Montpellier, France.

Legumes protect themselves from oxidative stresses and herbivores with a range of secondary compounds, including alkaloids, saponins and isoflavonoids. These have found antibiotic and health-promotive uses (Table 2). In some cases, analysis has not proceeded beyond a crude aqueous or solvent extract, but in many cases the specific active compound has been identified and tested. It is notable that the same compound can have several activities. The list presented in the table focuses on reports in refereed journals. Many of these reports also list species that were not affected by the treatment, but they are not included in Table 2 because of the space required.

Pueraria is a traditional Chinese medicinal plant (Gegen and Fengen in Chinese, kudzu in English), *Astragalus* species are medicinals in many cultures from Turkey to China (Huangqi in Chinese) and *Glycyrrhiza* species have been used as medicinals all over Eurasia (Gancao in Chinese, licorice in English). Modern science has tested and validated some of these properties and demonstrated their biochemical basis. Puerarin, the distinctive isoflavonoid glycoside of *Pueraria* has the glycoside C-linked rather than O-linked, so it does not get deglycosylated in the digestive tract, and it stays soluble unlike other isoflavones such as daidzein and genistein, accounting for some of the differences in its effectiveness (Cho et al. 2012). A comprehensive catalogue of the cardiovascular, cerebrovascular and other pharmacological activities of *Pueraria* and its components is provided by Wong et al. (2011). Licorice isoflavonoids and saponins have been found useful against oral and vaginal pathogens, confirming some of their traditional uses.

Anticancer activities of the indolizidine alkaloids swainsonine and castanospermine were identified in the 1990s (Molyneux et al. 2007), but there are few recent articles on these issues. Swainsonine is now known to be produced by endophytic fungi rather than the host plant (Braun et al. 2003, Cook et al. 2013).

The genus *Crotalaria* belongs to the Genisteae alliance clade (*Genisteae, Crotalarieae, Thermopsideae, Podylarieae, Liparieae,* and part of *Sophoreae*), characterized (unlike other legumes) by the production of alkaloids, generally quinolizidine alkaloids, except *Crotalaria* and some *Lotononis* spp. that produce pyrrolizidine ones. The remaining *Lotononis* spp. produce quinolizidine alkaloids. It appears that no legume is able to produce both types of alkaloids.

Numerous *Crotalaria* species are used as green manure all around the world. *Crotalaria* produce various active natural substances. More than 20 species are known for their

toxicity to ruminants (Watt & Breyer-Bradwijk, 1962; Verdcourt &Trump, 1969). Neal et al. (1935) purified an alkaloid (monocrotalin) from *C. spectabilis*. Later Kingsbury (1964) showed that monocrotaline was responsible for crotalism, a lethal illness affecting the nervous system, lung and liver. Since then, more than 50 pyrrolizidine alkaloids have been isolated from 45 *Crotalaria* species. (Mears & Mabry 1971; Kinghorn & Smolenski 1981; Polhill 1982). Further *Crotalaria* plants produce more biologically active molecules like flavonoids (Rao & Rao 1985; Yadava & Singh 1992; Wanjala & Majinda 1999) and chalcones (Yang et al. 1998; Kumar et al. 1999).

Alkaloid production is reputed to be the mechanism underlying the useful capacity of some Crotalaria species to control root-knot nematode populations (7 species), root-lesion nematodes Pratylenchus (2 species), Rotylenchulus (2 species), Helicotylenchus and Radopholus. Many field experiments in several tropical and sub-tropical countries have shown the resistance of Crotalaria to root-knot and other nematodes parasitizing crop plants, and that introduction of these resistant Crotalaria species into crop rotations resulted in significant decreases of some nematode populations in soils. In Brazil. Crotalaria spectabilis strongly reduced populations of *Meloidogyne incognita* (Huang et al. 1981). In India, Crotalaria juncea is recommended for management of Pratylenchus zeae in sugarcane plantations (Sundararraj & Mehta 1990). In Florida (USA), yields of eggplant and squash were enhanced after crop rotation with Crotalaria spectabilis had reduced Meloidogyne arenaria populations (McSorley et al. 1994). The development of Meloidogyne incognita larvae inoculated to Crotalaria spectabilis stopped at the 3rd stage (Sano et al. 1983). Aqueous extracts of Crotalaria spectabilis roots inhibited development of stage L2 larvae of *Meloidogyne incognita* (Subramaniyan & Vadivelu 1990).

Jourand et al. (2004) tested an extract of *Crotalaria grantiana* on tomato plants infested by *Meloidogyne incognita*. The biological activity was nematostatic: nematodes were not killed but were completely paralysed in a 1 mg/ml (w/v) extract, and the effect was reversible, as the paralysed juveniles recovered mobility in water and were able to infest a susceptible tomato plant. Freeze-dried aqueous extract from C. grantiana leaves added to a sterile sandy substrate at 1 mg/ml protected susceptible tomato plants from M. incognita infestation. This suggests a promising use of C. grantiana both as green manure and a natural alternative to synthetic chemicals in nematode population control, especially in integrated or organic pest management for vegetable crops of tropical and temperate areas. In a second step, these authors tested the sensitivity of 3 species of Meloidogyne to leaf and root extracts of 15 West African species of Crotalaria. In some species, both extracts were equally active, and in others, one was more active than the other, while one species had no detectable activity on the nematodes. Considering the greater biomass contribution of the leaves and stems compared to the roots when the plants are used as green manure, C. barkae, C. grantiana, C. pallida and C. podocarpa were considered the most efficient species, whatever the targeted species of *Meloidogyne*.

Agronomic trials in tropical or mediterranean conditions show that the use of *Crotalaria* plants as green manure can significantly reduce the dominant population of nematodes and effectively shield sensitive vegetable crops (Legume Futures project, in progress). Proliferation of nematodes in agricultural soils is a major problem worldwide, together with the high toxicity of the corresponding chemical treatments. Introducing *Crotalaria* plants having both capacities of symbiotic nitrogen fixation and nematode population control may be promising in agriculture (Wang et al., 2002, 2003).

CONCLUSIONS

F.L. Stoddard, Department of Agricultural Sciences, University of Helsinki, Finland

In addition to their vital role in sustainable agriculture, legumes have properties that make them valuable in animal feeding, industry, and medicine, in ways that are sometimes surprising and unexpected. Further uses are, no doubt, under development, and current uses can still be optimized by breeding, manufacture, or formulation.

Forage legumes, traditionally used only for feeding ruminants, have new roles as providers of leaf protein for feeding other animals, nitrogen for agroecosystems, and lignocellulose for biomass uses, in biorefining systems. Grain legumes can be used as whole-crop silage for ruminants given appropriate management, and protein fractions from them can be used as sustainable fish feeds, as long as the formulation takes care of amino acid, mineral and vitamin composition. Legumes have a vital role in bioremediation of oil-contaminated ground, where their independence of soil nitrogen is essential, but no greater role than any other plant in remediation of heavy metal contamination. The role of legumes in bioenergy production remains somewhat ambiguous. They have a clear role in providing nitrogen in some systems, but in many other situations, the nitrogen can come from methane digestate, or sewage sludge that cannot be used for food or feed production because of heavy metal or microbiological concerns.

Isoflavones, alkaloids, and other secondary compounds produced by legumes have proven to have a number of potential economic uses, including in the health industry. The effect of Crotalaria alkaloids, and some other legume chemicals, on nematodes has great implications for sustainable agricultural systems. Similarly, the suppression of the growth of malaria mosquitoes is an exciting outcome. The economics of some of these uses remain to be demonstrated, as do their effectiveness in the field rather than in the laboratory.

Table 2. Activities, source species, and effects of bioactive compounds from selected legumes.

Activity	Legume species	Target species	Active compounds	Reference
Antibacterial	Piptadeniastrum africanum	Staphylococcus aureus, Streptomyces mutans	Tannins	Brusotti et al. 2013
Antibacterial	Glycyrrhiza uralensis	Streptomyces mutans	Pterocarpene: Glycyrrhizol A; Isoflavonoid: 6,8 diisoprenyl-5,7,4'- trihydroxyisoflavone	He et al. 2006
Antibacterial	<i>Glycyrriza uralensis</i> and <i>G. glabra</i>	Streptomyces mutans, Porphyromonas gingivalis	Saponin: glycyrrhizin	Messier et al., 2012 (review)
Antibacterial	Glycyrriza uralensis	Streptomyces mutans, Porphyromonas gingivalis (in vitro)	Isoflavonoids: licoricidin, licorisoflavan A	Gafner et al. 2011
Antibacterial	Crotalaria pallida	Escherichia coli, Proteus sp., Ralstonia solanaceum, Erwinia sp. in vitro	Peptide <i>Cp</i> -AMP from seed protein	Pelegrini et al. 2009
Antibacterial	Ceratonia siliqua	Listeria monocytogenes in vitro	Total phenolics from leaves	Aissani et al. 2012
Antibacterial	Lupinus albus, L. angustifolius, L. hispanicus, L. luteus, L. mutabilis	Pseudomonas syringae, P. putida, Erwinia carotovora, in vitro	Quinolizidine alkaloids: lupinine	de la Vega et al. 1996
Antifungal	Piptadeniastrum africanum	Pyricularia grisea	Saponins	Brusotti et al. 2013
Antifungal	Glycyrriza uralensis and G.glabra	Candida albicans	Isoflavonoid: glabridin; Saponin: 18-β- glycyrrhetinic acid; Chalcone: licochalcone	Messier et al., 2012 (review)

Activity	Legume species	Target species	Active compounds	Reference
Antifungal	<i>Glycyrrhiza species</i> (probably <i>G.</i> <i>glabra</i>)	Candida albicans	Isoflavonoid: glabridin; Chalcone: licochalcone	Messier & Grenier 2011
Antifungal	Glycyrriza glabra	<i>Candida albicans</i> (in vitro)	Saponin: 18-β- glycyrrhetinic acid	Pellati et al. 2009
Antifungal	Glycyrriza glabra	<i>Candida albicans</i> drug-resistant mutants (<i>in vitro</i>), other filamentous fungi	lsoflavonoid: glabridin	Fatima et al. 2009
Antifungal	Crotalaria pallida	Fusarium oxysporum, Rhizoctonia solani in vitro	Peptide <i>Cp</i> -AMP from seed protein	Pelegrini et al. 2009
Antifungal	Astragalus verus	<i>Trichophytum</i> <i>verrucosum, in vitro</i> and <i>in vivo</i> on guinea pig	Aqueous extract	Mikaeili et al. 2012
Antifungal	Astragalus membranaceus	Trichoderma viride, Botrytis cinerea, Fusarium oxysporum, F. solani, in vitro	Chitinase	Kopparapu et al. 2011
Antifungal	Astragalus mongholicus (A. membranaceus var. mongholicus)	Botrytis cinerea, Fusarium oxysporum, Colletotrichum sp.	Galactose-recognizing lectin	Yan et al. 2005
Antifungal	Astragalus verrucosus	Aspergillus niger, Botrytis cinerea	Butanolic extract; saponin Astraverrucin II	Pistelli et al. 2002
Antifungal	Caesalpinia cacalaco	<i>Colletotrichum lindemuthianum</i> on common bean: fungistatic	Phenolics	Veloz-Garcia et al 2010
Antifungal	Prosopis alba, P. kuntzei, P. nigra, P. ruscifolia	<i>Coriolus versicolor</i> wood-rot fungus	Tannin: (-)-mesquitol	Pizzo et al. 2011

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Activity	Legume species	Target species	Active compounds	Reference
Anti-hypertensive	<i>Pueraria lobata</i> (in combination with <i>Salvia miltiorrhiza)</i>	Spontaneously hypertensive rat <i>in</i> <i>vivo</i> , aortic explant <i>in vitro</i>	Aqueous extract	Ng et al. 2011
Anti-inflammatory	Glycyrriza uralensis	Inflammatory cytokines IL-1 β , IL- 6, IL-8 and TNF- α	Isoflavonoids: licoricidin, licorisoflavan A	La et al. 2011 (cited by Messier et al. 2012)
Anti-inflammatory	Astragalus membranaceus	NO production from stimulated mouse macrophages	Isoflavonoid: Formononetin	Lai et al. 2013
Anti-insect	Dalbergia oliveri	<i>Aedes aegypti</i> , 3rd instar larvae and pupae, <i>in vivo</i>	Isoflavonoids: (+)- medicarpin, formononetin, (+)- violanone	Pluempanupat et al. 2013
Anti-insect	Sesbania grandiflora	Coptotermes formosanus	Methanolic extract of leaves	Elango et al. 2012
Anti-metabolic syndrome	Pueraria lobata	Spontaneously hypertensive rat <i>in</i> <i>vivo</i>	Isoflavones	Peng et al. 2009
Anti-metabolic syndrome	Glycyrrhiza glabra	Transactivation of peroxisome proliferator- activated receptor γ <i>in vitro</i> : antidiabetic	Total DMSO extract	Mueller & Jungbauer 2009
Anti-nematode	<i>Crotalaria</i> species	Meloidogyne incognita immobilization of adults and reduction of egg hatching; Rhabditis sp. repellence	Pyrrolizidine alkaloid Monocrotaline free base and N-oxide	Thoden et al. 2009
Anti-nematode	Crotalaria juncea	Various: also questions of human cirrhosis, genotoxicity, cancer & pulmonary hypertension	Dehydropyrrolizidine alkaloids	Colegate et al. 2012

Activity	Legume species	Target species	Active compounds	Reference
Anti-nematode	Crotalaria spectabilis, C. retusa, Mucuna pruriens	<i>Meloidogyne</i> spp., egg hatching & mobility reduced by exudates <i>in vitro</i> ; plants are non-host	Uncharacterized root exudates and plant extracts	Osei et al. 2010
Anti-nematode	Crotalaria juncea	<i>Meloidogyne</i> <i>incognita</i> suppressed, bacterivorous and fungivorous nematode populations promoted in field studies	Unidentified (presumed alkaloids)	Marahatta et al. 2010
Anti-nematode	Crotalaria juncea	Meloidogyne incognita reproduction reduced, Rotylenchus reniformis reproduction halted, pot and field studies	Unidentified (presumed alkaloids)	Robinson & Cook 2001
Anti-oxidant	Astragalus membranaceus and Glycyrrhiza uralensis	Free-radical scavenging assays and maintenance of viability of H ₂ O ₂ - challenged MRC-5 human fetal lung fibroblast cells	Phenolics and flavonoids in combination	Li et al. 2011
Anti-protozoa	Astragalus lentiginosus, A. mollissimus, other Astragalus spp., Oxytropis sericea, Swainsona spp.	<i>Trypanosoma cruzi</i> adhesion to host cells	Indolizidine alkaloid: swainsonine	James et al. 2004 (review)
Anti-protozoa	Castanospermum australe	<i>Plasmodium falciparum</i> adhesion to host cells	Indolizidine alkaloid: castanospermine	James et al. 2004 (review)
Anti-tumor	Glycyrrhiza species	Angiogenesis in mouse renal adenocarcinoma	Glycyrrhizic acid	Kim et al. 2013

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Activity	Legume species	Target species	Active compounds	Reference
Anti-tumorigenic	Astragalus membranaceus	Human colon cancer cells and gastric adeno- carcinoma cells <i>in</i> <i>vitro</i>	Total saponins	Auyeung et al. 2012
Antiviral	Castanospermum australe	Influenza virus, Human immunodeficiency virus, Dengue fever virus, by prevention of host-virus recognition	Indolizidine alkaloid: Castanospermine	Molyneux et al. 2007(review)
Hepato- protective, antioxidant	Astragalus corniculatus	Liver damage from CCl4 or paracetamol in rats	Purified saponin mixture	Vitcheva et al. 2013
Herbicide	Glycine max	Senna obtusifolia, Abutilon theophrasti in <i>Lolium perenne</i> turf	Fatty acid methyl ester, "biodiesel"	Vaughn & Holser 2007
Hypertensive	Glycyrrhiza species	Human kidney sodium metabolism	Glycyrrhizic acid, glycyrrhetinic acid	Miettinen et al. 2010
Hypertensive	Glycyrriza glabra	Increased expression of mineralocorticoid receptor in rat kidney	Glycyrrhizic acid	Ma et al. 2009
Hypoglycemic	<i>Lupinus</i> sp. (probably <i>L. albus</i>)	Mouse and human <i>in vivo</i>	Storage protein: γ- conglutin	Bertoglio et al. 2011
Insecticide	Pachyrhizus tuberosus	Inhibition of electron transport in animal and plant mitochondria	Rotenoids: rotenone, pachyrhizin and erosone from seeds	Ramos-de-la- Peña et al. 2013
Mammalian wound healing	Astragalus spp.	Human keratinocyte repair <i>in vitro</i> , rat skin <i>in vivo</i>	Cycloartane saponins	Sevimli-Gür et al. 2011

Legume-supported cropping systems for Europe

Activity	Legume species	Target species	Active compounds	Reference
Neurological disorders	Astragalus lentiginosus, A. mollissimus, other Astragalus spp., Oxytropis sericea, Swainsona spp.	Ruminants: α- mannosidase inhibition	Indolizidine alkaloid: swainsonine	James et al. 2004 (review)
Plant growth promotion (probably by change of soil microflora)	Lupinus exaltatus	<i>Capsicum annuum</i> , in pots	Total alkaloid extract, applied to soil	Przybylak et al. 2005
Pseudo- oestrogen	Pueraria lobata	Dyslipidemia and osteoporosis in mouse models of menopause	lsoflavone: puerarin	Cho et al. 2012
Teratogenesis	Lupinus formosus, L. arbustus, L. argenteus, L. sulphureus	Cattle and goats	Piperidine alkaloids	James et al. 2004 (review)
Teratogenesis	Lupinus albus, L. mutabilis	Cattle	Quinolizidine alkaloids	James et al. 2004 (review)

REFERENCES

- Aissani, N., Coroneo, V., Fattouch, S. & Caboni, P. 2012. Inhibitory effect of carob (*Ceratonia siliqua*) leaves methanolic extract on Listeria monocytogenes. Journal of Agricultural and Food Chemistry 60: 9954–9958.
- Arese, P., Bosia, A., Naitana, A., Gaetani, S., D'Aquino, M. & Gaetani, G.F. 1981. Effect of divicine and isouramil on red cell metabolism in normal and G6PD-deficient (Mediterranean variant) subjects. Possible role in the genesis of favism. Progress in Clinical Biological Research 55: 725–746.
- Aslaksen, M.A., Kraugerud, O.F., Penn, M., Svihus, B., Denstadli, V., Jörgensen, H.Y., Hillestad, M., Krogdahl, Å. & Storebakken, T. 2007. Screening of nutrient digestibilities and intestinal pathologies in Atlantic salmon, *Salmo salar*, fed diets with legumes, oilseeds, or cereals. Aquaculture 272: 541-555.
- Auyeung, K.K., Woo, P.K., Law, P.C. & Ko, J.K. 2012. *Astragalus* saponins modulate cell invasiveness and angiogenesis in human gastric adenocarcinoma cells. Journal of Ethnopharmacology 141: 635–641.
- Balogun, A.M. & Fetuga, B.L. 1986. Chemical composition of some underexploited leguminous crop seeds in Nigeria. Journal of Agricultural and Food Chemistry 34: 189-192.

BBC News. 2012. http://www.bbc.co.uk/news/uk-scotland-scotland-business-12169191

- Bertoglio, J.C., Calvo, M.A., Hancke, J.L., Burgos, R.A., Riva, A., Morazzoni, P., Ponzone, C., Magni, C. & Duranti, M. 2011. Hypoglycemic effect of lupin seed γ-conglutin in experimental animals and healthy human subjects. Fitoterapia 82: 933–938.
- Biswas, B., Scott, P.T. & Gresshoff, P.M. 2011. Tree legumes as feedstock for sustainable biofuel production: Opportunities and challenges. Journal of Plant Physiology 168: 1877–1884.
- Borreani, G., Chion, A.R., Colombini, S., Odoardi, M., Paoletti, R. & Tabacco, E. 2009. Fermentative profiles of field pea (*Pisum sativum*), faba bean (*Vicia faba*) and white lupin (*Lupinus albus*) silages as affected by wilting and inoculation. Animal Feed Science and Technology 151: 316-323.
- Boye, J., Zare. F. & Pletch, A. 2010. Pulse proteins: Processing, characterization, functional properties and applications in food and feed. Food Research International 43: 414–431.
- Braun, K., Romero, J., Liddell, C. & Creamer, R. 2003. Production of swainsonine by fungal endophytes of locoweed. Mycological Research 107: 980–988.
- Brehmer, B., Struik, P.C. & Sanders, J. 2008. Using an energetic and exergetic life cycle analysis to assess the best applications of legumes within a biobased economy. Biomass and Bioenergy 32: 1175-1186.
- Brinker, A. & Friedrich, C. 2012. Fish meal replacement by plant protein substitution and guar gum addition in trout feed. Part II: Effects on faeces stability and rheology. Biorheology 49: 27-48.
- Brunschwig, P. & Lamy, J.M. 2003. Les proteagineux contribuent a l'autonomie alimentaire du troupeau laitier alimente avec du mais ensilage, sans penaliser les performances. Fourrages 175: 395-402
- Brusotti, G., Tosi, S., Tava, A., Picco, A.M., Grisoli, P., Cesari, I. & Caccialanza, G. 2013. Antimicrobial and phytochemical properties of stem bark extracts from *Piptadeniastrum africanum* (Hook f.) Brenan. Industrial Crops and Products 43: 612-626.
- Butler, T.J., Muir, J.P., Huo, C. & Guretzky, J.A. 2013. Switchgrass biomass and nitrogen yield with overseeded cool-season forages in the southern Great Plains. Bioenergy Research 6: 44-52.
- Cho, H.J., Jun, H., Lee, J.H., Jia, Y., Hoang, M.H., Shim, J.-H., Park, K.-H. & Lee, S.-J. 2012. Acute effect of high-dose isoflavones from *Pueraria lobata* (Willd.) Ohwi on lipid and bone metabolism in ovariectomized mice. Phytotherapy Research 26: 1864–1871.
- Colegate, S.M., Gardner, D.R., Joy, R.J., Betz, J.M. & Panter, K.E. 2012. Dehydropyrrolizidine alkaloids, including monoesters with an unusual esterifying acid, from cultivated *Crotalaria juncea* (sunn hemp cv. 'Tropic Sun'). Journal of Agricultural and Food Chemistry 60: 3541–3550.
- Cook, D., Grum, D.S., Gardner, D.R., Welch, K.D. & Pfister, J.A. 2013. Influence of endophyte genotype on swainsonine concentrations in *Oxytropis sericea*. Toxicon 61: 105-111.
- Crépon, K., Marget, P., Peyronnet, C., Carroueé, B., Arese, P. & Duc, G. 2010. Nutritional value of faba bean (*Vicia faba* L.) seeds for feed and food. Field Crops Research 115: 329–339.

- D'Orazio, V., Ghanem, A. & Senesi, N. 2013. Phytoremediation of pyrene contaminated soils by different plant species. CLEAN Soil, Air, Water 41: 377-382.
- Davies, S.J. & Gouveia, A. 2010. Response of common carp fry fed diets containing a pea seed meal (*Pisum sativum*) subjected to different thermal processing methods. Aquaculture 305: 117–123.
- de la Vega, R., Gutierrez, M.P., Sanz, C., Calvo, R., Robredo, L.M., de la Cuadra, C. & Muzquiz, M. 1996. Bactericide-like effect of *Lupinus* alkaloids. Industrial Crops and Products 5: 141-148.
- De Silva, S.S. 2001. A global perspective of aquaculture in the new millennium. In 'Aquaculture in the Third Millennium. Technical Proceedings of the Conference on Aquaculture in the Third Millennium', Eds RP Subasinghe, P Bueno, MJ Phillips, C Hough, SE McGladdery, JR Arthur, Bangkok, Thailand, 20-25 February 2000. pp. 431-459. NACA, Bangkok and FAO, Rome.
- Dias, J., Alvarez, M.J., Arzel, J., Corraze, G., Diez, A., Bautista, J.M. & Kaushik, S.J. 2005. Dietary protein source affects lipid metabolism in the European seabass (*Dicentrarchus labrax*). Comparative Biochemistry and Physiology, Part A 142: 19-31.
- Diáz, M.J., Garcia, M.M., Eugenio, M.E., Tapias, R., Fernández, M. & López, F. 2007. Variations in fiber length and some pulp chemical properties of *Leucaena* varieties. Industrial Crops and Products 26: 142-150.
- El-Aafi, N., Brhada, F., Dary, M., Maltouf, A.F. & Pajuelo, E. 2012. Rhizostabilization of metals in soils using *Lupinus luteus* inoculated with the metal resistant rhizobacterium *Serratia* sp. MSMC541. International Journal of Phytoremediation 14: 261-274.
- Elango, G., Rahuman, A.A., Kamaraj, C., Bagavan, A., Zahir, A.A., Santhoshkumar, T., Marimuthu, S., Velayutham, K., Jayaseelan, C., Kirthi, A.V. & Rajakumar, G. 2012. Efficacy of medicinal plant extracts against Formosan subterranean termite, *Coptotermes formosanus*. Industrial Crops and Products 36: 524–530.
- Eriksson, T. 2010. Nitrogen metabolism in dairy cows fed restricted amounts of grass-clover silage supplemented with seeds from narrow-leafed lupin or pea. Livestock Science 131: 39-44.
- Fatima, A., Gupta, V.K., Luqman, S., Negi, A.S., Kumar, J.K., Shanker, K., Saikia, D., Srivastava, S., Darokar, M.P. & Khanuja, S.P.S. 2009. Antifungal activity of *Glycyrrhiza glabra* extracts and its active constituent glabridin. Phytotherapy Research 23: 1190-1193.
- Foresight. 2011. The Future of Food and Farming. Final Project Report. The Government Office for Science, London, UK.
- Fraser, M.D., Fychan, R. & Jones, R. 2005a. The effect of harvest date and inoculation on the yield and fermentation characteristics of two varieties of white lupin (Lupinus albus) when ensiled as a whole-crop. Animal Feed Science and Technology 119:307-322.
- Fraser, M.D., Fychan, R. & Jones, R. 2005ab. Comparative yield and chemical composition of two varieties of narrow-leafed lupin (*Lupinus angustifolius*) when harvested as whole-crop, moist grain and dry grain. Animal Feed Science and Technology 120: 43-50.
- Froidmont, E. & Bartiaux-Thill, N. 2004. Suitability of lupin and pea seeds as a substitute for soybean meal in high-producing dairy cow feed. Animal Research 53: 475–487.
- Fru-Nji, F., Niess, E. & Pfeffer, E. 2007. Effect of graded replacement of soyabean meal by faba beans (*Vicia faba* L.) or field peas (*Pisum sativum* L.) in rations for laying hens on egg production and quality. Journal of Poultry Science 44: 34-41.
- Garner, S., Bergeron, C., Villinski, J.R., Godejohann, M., Kessler, P., Cardelina, J.H.II, Ferreira, D., Feghali,
 K. & Grenier, D. 2011. Isoflavonoids and coumarins from *Glycyrrhiza uralensis*: antibacterial activity against oral pathogens and conversion of isoflavans into isoflavan-quinones during purification. Journal of Natural Products 74: 2514-2519.
- Gatlin, D.M., Barrows, F.T., Brown, P., Dabrowski, K., Gibson Gaylord, T., Hardy, R.W., Herman, E., Hu, G., Krogdahl, A., Nelson, R., Overturf, K., Rust, M., Sealey, W., Skonberg, D., Souza, E.J., Stone, D., Wilson, R. & Wurtele, E. 2007. Expanding the utilization of sustainable plant products in aquafeeds: a review. Aquaculture Research 38: 551–579.

- Glencross, B.D., Boujard, T. & Kaushik, S.J. 2003. Influence of oligosaccharides on the digestibility of lupin meals when fed to rainbow trout (*Oncorhynchus mykiss*). Aquaculture 219: 703-713.
- Glencross, B., Hawkins, W., Evans, D., Rutherford, N., Dods, K., McCafferty, P. & Sipsas, S. 2008. Evaluation of the influence of *Lupinus angustifolius* kernel meal on dietary nutrient and energy utilization efficiency by rainbow trout (*Oncorhynchus mykiss*). Aquaculture Nutrition 14: 129-138.
- Gomes, E.F., Rema, P. & Kaushik, S.J. 1995. Replacement of fish meal by plant proteins in the diet of rainbow trout (*Oncorhynchus mykiss*): digestibility and growth performance. Aquaculture 130: 177-186.
- González-García, S., Moreira, M.T. & Feijoo, G. 2010. Environmental performance of lignocellulosic bioethanol production from alfalfa stems. Biofuels, Bioproducts and Biorefining 4: 118-131.
- González-García, S., Moreira, M.T., Feijoo, G. & Murphy, R.J. 2012. Comparative life cycle assessment of ethanol production from fast-growing wood crops (black locust, eucalyptus and poplar). Biomass and Bioenergy 39: 378-388.
- Griffiths, D.W. & Jones, D.I.H. 1977. Cellulase inhibition by tannins in the testa of field beans (*Vicia faba*). Journal of the Science of Food Agriculture 28: 983-989.
- Griffiths, D.W. 1986. The inhibition of digestive enzymes by polyphenolic compounds. Advances in Experimental Medicine and Biology 199: 509.
- Grünewald, H., Böhm, C., Quinkenstein, A., Grundmann, P., Eberts, J. & Wühlisch, G. von. 2009. *Robinia pseudoacacia* L.: A lesser known tree species for biomass production. BioEnergy Research 2: 123-133.
- Gunawardena, C.K., Zijlstra, R.T., Goonewardene, L.A. & Beltranena, E. 2010. Protein and starch concentrates of air-classified field pea and zero-tannin faba bean for weaned pigs. Journal of Animal Science 88: 2627-2636.
- He, J., Chen, L., Hever, D., Shi, W. & Lu, Q.-Y. 2006. Antibacterial compounds from *Glycyrrhiza uralensis*. Journal of Natural Products 69: 121-124.
- Hill, J., Nelson, E., Tilman, D., Polasky, S. & Tiffany, D. 2006. Environmental, economic, and energetic costs and benefits of biodiesel and ethanol biofuels. Proceedings of the National Academy of Sciences of the USA 103: 11207-11211.
- Hosamani, K.M., Patil, A.S. & Pattanashettar, R.S. 2002. *Acacia arabica* varieties -- *Telia babul*, *Vediana* and *Cupressiformis* seed oils: a moderate source of coronaric and cyclopropene fatty acids. Industrial Crops and Products 15: 131-137.
- Hosamani, K.M., Patil, A.S. & Pattanashettar, R.S. 2002. *Acacia arabica* varieties Telia babul, Vediana and Cupressiformis seed oils: a moderate source of coronaric and cyclopropene fatty acids. Industrial Crops and Products 15: 131-137.
- Howieson, J.G., Yates, R.J., Foster, K.J., Real, D. & Besier, R.B. 2008. Prospects for the future use of legumes. Chapter 12 in: Nitrogen-fixing Leguminous Symbioses, eds Dilworth, M.J., James, E.K., Sprent, J.I. & Newton, W.E. Berlin, Germany: Springer.
- Huang, C.S., Tenante, R.C.V., Silva, F.C.C. & Da Lara, J.A.R. 1981. Effect of *Crotalaria spectabilis* and two nematicides, on numbers of *Meloidogyne incognita* and *Helicotylenchus dihystera*. Nematologica 27: 1-5.
- Ingle, N.P. & Doke, S.S. 2006. Analysis of sunnhemp fibers processed using jute spinning system. Industrial Crops and Products 23: 235–243.
- Ishida, K., Yani, S., Kitagawa, M., Oishi, K., Hirooka, H. & Kumagai, H. 2012. Effects of adding food byproducts mainly including noodle waste to total mixed ration silage on fermentation quality, feed intake, digestibility, nitrogen utilization and ruminal fermentation in wethers. Animal Science Journal 81: 735-742.
- James, L.F., Panter, K.E., Gaffield, W. & Molyneux, R.J. 2004. Biomedical applications of poisonous plant research. Journal of Agricultural and Food Chemistry 52: 3211-3230.
- Jensen, E.S., Peoples, M.B., Boddey, R.M., Gresshoff, P.M., Hauggaard-Nielsen, H., Alves, B.J.R. & Morrison, M.J. 2012. Legumes for mitigation of climate change and the provision of feedstock for biofuels and biorefineries. A review. Agronomy for Sustainable Development 32: 329-364.

- Jezierny, D., Mosenthin, R., Sauer, N., Roth, S., Piepho, H.P., Rademacher, M. & Eklund, M. 2011. Chemical composition and standardised ileal digestibilities of crude protein and amino acids in grain legumes for growing pigs. Livestock Science 138: 229-243.
- Jourand, P., Rapior, S., Fargette, M. & Mateille, T. 2004. Nematostatic activity of aqueous extracts of West African *Crotalaria* species. Nematology 6: 765-771
- Jussila, M.M., Jurgens, G., Lindström, K. & Suominen, L. 2006. Genetic diversity of culturable bacteria in oil-contaminated rhizosphere of *Galega orientalis*. Environmental Pollution 139: 244-257.
- Kaksonen, A.H., Jussila, M.M., Lindström, K. & Suominen, L. 2006. Rhizosphere effect of *Galega orientalis* in oil-contaminated soil. Soil Biology and Biochemistry 38: 817-827.
- Kamm, B., Hille, C., Schönicke, P. & Dautzenberg, G. 2010. Green biorefi nery demonstration plant in Havelland (Germany). Biofuels, Bioproducts and Biorefining 4: 253-262.
- Karababa, E. & Coskuner, Y. 2013. Physical properties of carob bean (Ceratonia siliqua L.): An industrial gum yielding crop. Industrial Crops and Products 42: 440– 446.
- Kaushik, S.J., Covès, D., Dutto, G. & Blanc, D. 2004. Almost total replacement of fish meal by plant protein sources in the diet of a marine teleost, the European seabass, *Dicetrarchus labrax*. Aquaculture 230: 391-404.
- Kawasaki, A., Watson, E.R. & Kertesz, M.A. 2012. Indirect effects of polycyclic aromatic hydrocarbon contamination on microbial communities in legume and grass rhizospheres. Plant and Soil 358:169-182.
- Kim, K.-J., Choi, C.-S., Kim, K.-W. & Jeong, J.-W. 2013. The anti-angiogenic activities of glycyrrhizic acid in tumor progression. Phytotherapy Research 27: 841–846.
- King, R.D. & Dietz, H.M. 1987. Air classification of rapeseed meal. Cereal Chemistry 64: 411-413.
- Kinghorn, A.D. & Smolenski, S.J. 1981. Alkaloids in Papilionoideae. P585-589. In: Advances in Legume Systematics, eds Polhill, R.M. & Raven, P.H. Royal Botanic Gardens, Kew, England.
- Kingsbury, J.M. 1964. Poisonous plants of the United States and Canada. Prentice Hall, New York, USA.
- Kirkpatrick, W.D., White, P.M. Jr, Wolf, D.C., Thoma, G.J. & Reynolds C.M. 2006. Selecting plants and nitrogen rates to vegetate crude-oil--contaminated soil. International Journal of Phytoremediation 8: 285-297.
- Kopparapua, N.K., Liub, Z., Fei, F., Yan, Q. & Jiang, Z. 2011. Purification and characterization of a chitinase (sAMC) with antifungal activity from seeds of *Astragalus membranaceus*. Process Biochemistry 46: 1370–1374.
- Krogdahl, Å., Bakke-McKellep, A.M. & Baeverfjord, G. 2003. Effects of graded levels of standard soybean meal on intestinal structure, mucosal enzyme activities, and pancreatic response in Atlantic salmon (*Salmo salar* L.). Aquaculture Nutrition <u>9</u>: 361–371.
- Kumar, J.K., Narender, T., Rao, M.S., Rao, P.S. & Toth, G. 1999. Further Dihydrochalcones from *Crotalaria ramosissima*. Journal of the Brazilian Chemical Society 10: 278-280.
- Lai, P.K.-K., Chan, J.Y.-W., Cheng, L., Lau, C.-P., Han, S.Q.-B., Leung, P.-C., Fung, K.-P. & Lau, C.B.-S. 2013. Isolation of anti-inflammatory fractions and compounds from the root of *Astragalus membranaceus*. Phytotherapy Research 27: 581–587.
- Laudadio, V. & Tufarelli V. 2011. Dehulled-micronised lupin (Lupinus albus L. cv. Multitalia) as the main protein source for broilers: influence on growth performance, carcass traits and meat fatty acid composition. Journal of the Science of Food and Agriculture 91: 2081–2087.
- Laudadio, V., Ceci E. & Tufarelli V. 2011. Productive traits and meat fatty acid profile of broiler chickens fed diets containing micronized fava beans (*Vicia faba* L. var. *minor*) as the main protein source. Journal of Applied Poultry Research 20:12-20.
- Létang, C., Samson, M.F., Lasserre, T.M., Chaurand, M. & Abécassis, J. 2002. Production of starch with very low protein content from soft and hard wheat flours by jet milling and air classification. Cereal Chemistry 79: 535-543.
- Li, M., Xua, Y., Yang, W., Li, J., Xu, X., Zhang, X., Chen, F. & Li, D. In vitro synergistic anti-oxidant activities of solvent-extracted fractions from *Astragalus membranaceus* and *Glycyrrhiza uralensis*. LWT Food Science and Technology 44: 1745-1751.

- Lindström, K., Jussila, M.M., Hintsa, H., Kaksonen, A., Mokelke, L., Mäkeläinen, K., Pitkäjärvi, J. & Suominen, L. 2003. Potential of the *Galega Rhizobium galegae* system for bioremediation of oil-contaminated soil. Food Technology and Biotechnology 41: 11-16.
- Ma, D.K., Bae, E.H., Kim, I.J., Choi, K.C., Kim, S.H., Lee, J.U. & Kim, S.W. 2009. Increased renal expression of nitric oxide synthase and atrial natriuretic peptide in rats with glycyrrhizic-acid-induced hypertension. Phytotherapy Research 23: 206-211.
- Marahatta, S.P., Wang, K.-H., Sipes, B.S. & Hooks, C.R.R. 2010. Strip-tilled cover cropping for managing nematodes, soil mesoarthropods, and weeds in a bitter melon agroecosystem. Journal of Nematology 42: 111–119.
- Mariotti, M., Masoni, A., Ercoli, L. & Arduini, I. 2012. Optimizing forage yield of durum wheat/field bean intercropping through N fertilization and row ratio. Grass and Forage Science 67:243-254.
- Marley, C., Davies, D., Fisher, B., Fychan, R., Sanderson, R., Jones, R. & Abberton, M. 2008. Effects of incorporating yellow lupins into concentrate diets compared with soya on milk production and milk composition when offered to dairy cows. Lupins for health and wealth: Proceedings of the 12th International Lupin Conference, Fremantle, Western Australia, 14-18 September 2008. pp. 115-117.
- Mazurkiewicz, J. 2009. Utilization of domestic plant components in diets for common carp (*Cyprinus carpio* L.). Archives of Polish Fisheries 17: 5-39.
- McSorley, R., Dickson, D.W., Brito, J.A., De Hewlett, T.E. & Frederick, J.J. 1994. Effects of tropical rotation crops on *Meloidogyne arenaria* population densities and vegetable yields in microplots, Journal of Nematology, 26:175-181.
- Mears, J.A. & Mabry, T.J. 1971. Alkaloids in Leguminosae. Pp 73-178. In : Chemotaxonomy of the Leguminosae, eds Harborn, J.B., Boulter, D. & Turner, B.L. Academic Press, London, England.
- Messier, C. & Grenier, D. 2011. Effect of licorice compounds licochalcone A, glabridin and glycyrrhizic acid on growth and virulence properties of Candida albicans. Mycoses 54: e801–e806.
- Messier, C., Epifano, F., Genovese, S. & Grenier, D. 2012. Licorice and its potential beneficial effects in common oro-dental diseases. Oral Diseases 18: 32-39.
- Miettinen, H.E., Piippo, K., Hannila-Handelberg, T., Paukku, K., Hiltunen, T.P., Gautschi, I., Schild, L. & Kontula, K. 2010. Licorice-induced hypertension and common variants of genes regulating renal sodium reabsorption. Annals of Medicine 42: 465–474.
- Mikaeili, A., Modaresi, M., Karimi, I., Ghavimi, H., Fathi, M. & Jalilian, N. 2012. Antifungal activities of *Astragalus verus* Olivier. against *Trichophyton verrucosum* on *in vitro* and *in vivo* guinea pig model of dermatophytosis. Mycoses 55: 318–325.
- Miller, M.R., Nichols, P.D. & Carter, C.G. 2008. n-3 Oil sources for use in aquaculture alternatives to the unsustainable harvest of wild fish. Nutrition Research Reviews 21: 85–96.
- Molina-Poveda, C., Lucas, M. & Jover, M. 2013. Evaluation of the potential of Andean lupin meal (*Lupinus mutabilis* Sweet) as an alternative to fish meal in juvenile *Litopenaeus vannamei* diets. Aquaculture 410/411: 148-156.
- Molyneux, R.J., Lee, S.T., Gardner, D.R. Panter, K.E. & James, L.F. 2007. Phytochemicals: the good, the bad and the ugly? Phytochemistry 68: 2973-2985.
- Morris, J.B. 1997. Bio-functional legumes with nutraceutical, pharmaceutical and industrial uses. Economic Botany 57: 254-261.
- Morris, J.B. 2003. Special-purpose legume genetic resources conserved for agricultural, industrial and pharmaceutical use. Economic Botany 51: 251-263.
- Moss, A., Allison, R., Stroud, A. & Collins, C. 2000. Evaluation of heat-treated lupins, beans and rapeseed meal as protein sources for dairy cows. HGCA Project Report OS45: 45 pp.
- Mueller, M. & Jungbauer, A. 2009. Culinary plants, herbs and spices A rich source of $PPAR_{\chi}$ ligands. Food Chemistry 117: 660-667.
- National Research Council. 2011. Nutrient Requirements of Fish and Shrimp. National Academies Press, Washington DC, USA.

- Naylor, R.L., Hardy, R.W., Bureau, D.P., Chiu, A., Elliott, M., Farrell, A.P., Forster, I., Gatlin, D.M., Goldburg, R.J., Hua, K. & Nichols, P.D. 2009. Feeding aquaculture in an era of finite resources. Proceedings of the National Academy of Sciences of the USA 106: 15103-15110.
- Neal, W.N., Rusoff, L.L. & Ahmann, C.F. 1935. The isolation and some properties of an alkaloid from *Crotalaria spectabilis* Roth. Journal of the American Chemical Society 57: 2560-2561.
- Nehdi, I.A., Sbihi, H., Tan, C.P., Zarrouk, H., Khalil, M.I. & Al-Resayes, S.I. 2012. Characteristics, composition and thermal stability of *Acacia senegal* (L.) Willd. seed oil. Industrial Crops and Products 36: 54-58.
- Ng, C.F., Koon, C.M., Cheung, D.W.S., Lam, M.Y., Leung, P.C., Lau, C.B.S. & Fung, K.P. 2011. The antihypertensive effect of Danshen (*Salvia miltiorrhiza*) and Gegen (*Pueraria lobata*) formula in rats and its underlying mechanisms of vasorelaxation. Journal of Ethnopharmacology 137: 1366–1372.
- Niwinska, B. & Andrzejewski, M. 2011. Effect of the type of non-fibre carbohydrates in grass silage-based diets on *in sacco* ruminal degradability and protein value of lupin (*Lupinus angustifolius* L. cv. Sonet) seeds ground to different particle sizes. Czech Journal of Animal Sciences 56: 231–241.
- Nonnoi, F., Chinnaswamy, A., Garcia la Torre, V.S. de, Coba de la Pena, T., Lucas, M.M. & Pueyo, J.J. 2012. Metal tolerance of rhizobial strains isolated from nodules of herbaceous legumes (*Medicago* spp. and *Trifolium* spp.) growing in mercury-contaminated soils. Applied Soil Ecology 61: 49-59.
- Olkowski, B.I., Janiuk, I. & Jakubczak, A. 2010. Effect of enzyme preparation with activity directed towards degradation of non starch polysaccharides on yellow lupine seed based diet for young broilers. Acta Veterinaria Brno 79: 395-402.
- Olsen, H.S. & Andersen, J.H. 1978. The estimation of vicine and convicine in faba beans (Vicia faba L.) and isolated fababean proteins. Journal of the Science of Food and Agriculture 29: 323-331.
- Osei, K., Gowen, S.R., Pembroke, B., Brandenburg, R.L. & Jordan, D.L. 2010. Potential of leguminous cover crops in management of a mixed population of root-knot nematodes (*Meloidogyne* spp.). Journal of Nematology 42: 173–178.
- Pakarinen, A., Maijala, P., Jaakkola, S., Stoddard, F.L., Kymalainen, M. & Viikari, L. 2011. Evaluation of preservation methods for improving biogas production and enzymatic conversion yields of annual crops. Biotechnology for Biofuels 4: art. 20.
- Palander, S., Laurinen, P., Perttila, S., Valaja, J. & Partanen, K. 2006. Protein and amino acid digestibility and metabolizable energy value of pea (*Pisum sativum*), faba bean (*Vicia faba*) and lupin (*Lupinus angustifolius*) seeds for turkeys of different age. Animal Feed Science and Technology 127: 89-100.
- Pelegrini, P.B., Farias, L.R., Saude, A.C.M., Costa, F.T., Block, C. Jr., Silva, L.P., Oliveira, A.S., Gomes, C.E.M., Sales, M.P. & Franco, O.L. 2009. A novel antimicrobial peptide from *Crotalaria pallida* seeds with activity against human and phytopathogens. Current Microbiology 59: 400–404.
- Pelgrom, P.J.M., Vissers, A.M., Boom, R.M. & Schutyser, M.A.I. 2013. Dry fractionation for production of functional pea protein concentrates. Food Research International 53: 232–239.
- Pellatti, D., Fiore, C., Armanini, D., Rassu, M. & Bertoloni, G. 2009. *In vitro* effects of glycyrrhetinic acid on the growth of clinical isolates of *Candida albicans*. Phytotherapy Research 23: 572–574.
- Peng, N., Prasain, J.K., Dai, Y., Moore, R., Arabshahi, A., Barnes, S., Carlson, S. & Wyss, J.M. 2009. Chronic dietary kudzu isoflavones improve components of metabolic syndrome in stroke-prone spontaneously hypertensive rats. Journal of Agricultural and Food Chemistry 57: 7268-7273.
- Pistelli, L., Bertoli, A., Lepori, E., Morelli, I. & Panizzi, L. 2002. Antimicrobial and antifungal activity of crude extracts and isolated saponins from *Astragalus verrucosus*. Fitoterapia 73: 336-339.
- Pizzo, B., Pometti, C.L., Charpentier, J.-P., Boizot, N. & Saidman, B.O. 2011. Relationships involving several types of extractives of five native argentine wood species of genera *Prosopis* and *Acacia*. Industrial Crops and Products 34: 851-859.
- Pluempanupat, S., Kumrungsee, N., Pluempanupat, W., Ngamkitpinyo, K., Chavasiri, W., Bullangpoti, V. & Koul, O. 2013. Laboratory evaluation of *Dalbergia oliveri* (Fabaceae: Fabales) extracts and isolated isoflavonoids on *Aedes aegypti* (Diptera: Culicidae) mosquitoes. Industrial Crops and Products 44: 653–658.

Polhill, R.M. 1982. Crotalaria in Africa and Madagascar. Balkema, Rotterdam, The Netherlands. 389p.

- Prajapati, V.D., Jani, G.K., Moradiya, N.G., Randeria, N.P., Nagar, B.J. 2013. Locust bean gum: a versatile biopolymer. Carbohydrate Polymers 94: 814-821.
- Przybylak, J., Ciesiołka, D., Wysocka, W., Garcia-Lopez, P.M., Ruiz-Lopez, M.A., Wysocki, W. & Gulewicz, K. 2005. Alkaloid profiles of Mexican wild lupin and an effect of alkaloid preparation from *Lupinus exaltatus* seeds on growth and yield of paprika (*Capsicum annuum* L.). Industrial Crops and Products 21: 1–7.
- Radwan, S.S., Al-Awadhi, H. & El-Nemr, I.M. 2000. Cropping as a phytoremediation practice for oily desert soil with reference to crop safety as food. International Journal of Phytoremediation 2: 383-396.
- Ramos-de-la-Peña, A.M., Renard, C.M.G.C., Wicker, L. & Contreras-Esquivel, J.C. 2013. Advances and perspectives of *Pachyrhizus* spp. in food science and biotechnology. Trends in Food Science & Technology 29: 44-54.
- Rao, G.V. & Rao, P.S. 1985. A new flavonol glycoside from flowers of *Crotalaria verrucosa*. Fitoterapia 56:175-177.
- Robinson, A.F. & Cook, C.G. 2001. Root-knot and reniform nematode reproduction on kenaf and sunn hemp compared with that on nematode resistant and susceptible cotton. Industrial Crops and Products 13: 249–264.
- Scott, P.T., Pregelj, L., Ning, C., Hadler, J.S., Djordjevic, M.A. & Gresshoff, P.M. 2008. *Pongamia pinnata:* an untapped resource for the biofuels industry of the future. BioEnergy Research 1: 2-11.
- Scottish Salmon Producers' Organisation. 2012. Scottish Salmon Farming Industry Research Report. Scottish Salmon Producers' Limited, Perth, UK.
- Serrano, J.E. 1989. Chemical and nutritive values of three ensiled residues (broad beans, peas and soyabean), in comparison with yellow lupin silage. Proceedings of the XVI International Grassland Congress, 4-11 October 1989, Nice, France, pp. 983-984.
- Sevimli-Gür, C., Onbasılar, I., Atilla, P., Genc, R., Cakar, N., Deliloglu-Gürhana, I. & Bedir, E. 2011. In vitro growth stimulatory and in vivo wound healing studies on cycloartane-type saponins of *Astragalus* genus. Journal of Ethnopharmacology 134: 844–850.
- Shahsavari, E., Adetutu, E.M., Anderson, P.A. & Ball, A.S. 2013. Necrophytoremediation of phenanthrene and pyrene in contaminated soil. Journal of Environmental Management 122: 105-112.
- Shapiro, M. & Galperin, V. 2005. Air classification of solid particles: a review. Chemical Engineering and Processing: Process Intensification 44: 279-285.
- Sibakov, J., Myllymäki, O., Holopainen, U., Kaukovirta-Norja, A., Hietaniemi, V., Pihlava, J.M., Poutanen, K. & Lehtinen, P. 2010. Lipid removal enhances separation of oat grain cell wall material from starch and protein. Journal of Cereal Science 54: 104-109.
- Smith, D.M., Tabrett, S.J. & Glencross, B. D. 2007b. Growth response of the black tiger shrimp, *Penaeus monodon* fed diets containing different lupin cultivars. Aquaculture 269: 436-446.
- Smith, D.M., Tabrett, S.J., Irvin, S.J., Wakeling, J., Glencross, B.D. & Harris, D. 2007a. Response of the black tiger shrimp, *Penaeus monodon* to feed containing the lupin alkaloid, gramine. Aquaculture 272: 556-563.
- Smith, L.A., Houdijk, J.G.M., Homer, D. & Kyriazakis, I. 2013. Effects of dietary inclusion of pea and faba bean as a replacement for soybean meal on grower and finisher pig performance and carcass quality. Journal of Animal Science 91: 3733-3741.
- Steenfeldt, S., Gonzalez, E. & Knudsen, K.E.B. 2003. Effects of inclusion with blue lupins (*Lupinus angustifolius*) in broiler diets and enzyme supplementation on production performance, digestibility and dietary AME content. Animal Feed Science and Technology 110: 185-200.
- Stoddard, F.L. 2008. Bioenergy, legumes and biotechnology. Recent Advances in Biotechnology, eds. B.N. Prasad and L. Mathew. Excel India Publishers, New Delhi, India, pp. 63-66.
- Subramaniyan, S. & Vadivelu, S. 1990. Effects of *Crotalaria spectabilis* extracts on *Meloidogyne incognita*. International Nematology Network Newsletter, 7: 8-9.

- Sun, M., Fu, D., Teng, Y., Shen, Y., Luo, Y., Li, Z. & Christie, P. 2011. In situ phytoremediation of PAHcontaminated soil by intercropping alfalfa (*Medicago sativa* L.) with tall fescue (*Festuca arundinacea* Schreb.) and associated soil microbial activity. Journal of Soils and Sediments 11: 980-989.
- Sundararraj, P. & Mehta, U.K. 1990. Host status of some economic crops to *Pratylenchus zeae* and their influence on subsequent sugarcane crops. Indian Journal of Nematology, 26: 175-181.
- Suominen, L., Jussila, M.M., Mäkeläinen, K., Romantschuk, M. & Lindström, K. 2000. Evaluation of the *Galega Rhizobium galegae* system for the bioremediation of oil-contaminated soil. Environmental Pollution 107: 239-244.
- Swick, R.A. & Cremer, M.C. 2001. Livestock production: a model for aquaculture?, Guest Lecture. In 'Aquaculture in the Third Millennium. Technical Proceedings of the Conference on Aquaculture in the Third Millennium', Eds RP Subasinghe, P Bueno, MJ Phillips, C Hough, SE McGladdery, JR Arthur, Bangkok, Thailand, 20-25 February 2000. pp.49-60. NACA, Bangkok and FAO, Rome.
- Tacon, A.G.J. 1997. A review of the state of world aquaculture. FAO Fisheries Department, FAO Fisheries Circular No. 886 FIRI/C886 (Rev.1), Rome, ISSN 0429-9329.
- Thoden, T.C., Boppré, M. & Hallmann, J. 2009. Effects of pyrrolizidine alkaloids on the performance of plant-parasitic and free-living nematodes. Pest Management Science 65: 823–830.
- Thomsen, M.H. & Hauggaard-Nielsen, H. 2008. Sustainable bioethanol production combining biorefinery principles using combined raw materials from wheat undersown with clover-grass. Journal of Industrial Microbiology & Biotechnology 35: 303-311.
- Trushinski, J.T., Kasper, C.S. & Kohler, C.C. 2006. Challenge and opportunities in finfish nutrition. North American Journal of Aquaculture 68: 122-140.
- Tyler, R.T. & Panchuk, B.D. 1982. Effect of seed moisture content on the air classification of field peas and faba beans. Cereal Chemistry 59: 31-33.
- Tyler, R.T., Youngs, C.G. & Sosulski, F.W. 1981 Air classification of legumes. I. Separation efficiency, yield, and composition of the starch and protein fractions. Cereal Chemistry 58: 144 148.
- Tzanakakis, V.A., Chatzakis, M.K. & Angelakis, A.N. 2012. Energetic environmental and economic assessment of three tree species and one herbaceous crop irrigated with primary treated sewage effluent. Biomass and Bioenergy 47: 115-124.
- United Nations. 2009. World population prospects: the 2008 Revision. Population Division of the Department of Economic and Social Affairs of the United Nations Secretariat, June.
- Varadi, L., Szucs, I., Pekar, F., Blokhin, S. & Csavas, I. 2001 Aquaculture development trends in Europe. In 'Aquaculture in the Third Millennium. Technical Proceedings of the Conference on Aquaculture in the Third Millennium', Eds RP Subasinghe, P Bueno, MJ Phillips, C Hough, SE McGladdery, JR Arthur, Bangkok, Thailand, 20-25 February 2000. pp. 397-416. NACA, Bangkok and FAO, Rome.
- Vasanthan, T. & Bhatty, R.S. 1995. Starch purification after pin milling and air classification of waxy, normal, and high amylose barleys. Cereal Chemistry 72: 379-384.
- Vaughn, S.E. & Holser, R.A. 2007. Evaluation of biodiesels from several oilseed sources as environmental friendly contact herbicides. Industrial Crops and Products 26: 63-68.
- Verdcourt, B. & Trump, E.C. 1969. Common poisonous plants of East Africa. Collins, London, England.
- Vilarino, M., Metayer, J.P., Crepon, K. & Duc, G. 2009. Effects of varying vicine, convicine and tannin contents of faba bean seeds (*Vicia faba* L.) on nutritional values for broiler chicken. Animal Feed Science and Technology 150:114-121.
- Vinay, B.J. & Sindhu Kanya, T.C. 2008. Effect of detoxification on the functional and nutritional quality of proteins of karanja seed meal. Food Chemistry 106: 77-84.
- Vismaya, Eipeson, W.S., Manjunatha, J.R., Srinivas, P. & Sindhu Kanya, T.C. 2010. Extraction and recovery of karanjin: a value addition to karanja (*Pongamia pinnata*) seed oil. Industrial Crops and Products 32: 118-122.
- Vitcheva, V., Simeonova, R., Krasteva, I., Nikolov, S. & Mitcheva M. 2013. Protective effects of a purified saponin mixture from *Astragalus corniculatus* Bieb., *in vivo* hepatotoxicity models. Phytotherapy Research 27: 731–736.

- Wang, D., Lebauer, D.S. & Dietze, M.C. 2010. A quantitative review comparing the yield of switchgrass in monocultures and mixtures in relation to climate and management factors. Global Change Biology Bioenergy 2: 16-25.
- Wang, K.H., McSorley, R. & Gallaher, R.N. 2003. Effect of *Crotalaria juncea* as amendment on nematode communities in soil with different agricultural histories. J. Nematology 35: 294-301.
- Wang, K.H., Sipes, B.S. & Schmitt, D.P. 2002. *Crotalaria* as cover crop for nematode management : a review. Nematropica 32: 35-57.
- Wanjala, C.C.W. & Majinda, R.R.T. 1999. Flavonoids glycosides from *Crotalaria podocarpa*. Phytochemistry 51: 705-707.
- Watt, J.M. & Breyer-Brandwijk, M.G. 1962. The Medicinal & Poisonous Plants of Southern & Eastern Africa. In: Being an account of their medicinal and other uses, chemical composition, pharmacological effects and toxicology in man and animal, 2nd edn. E&S Livingstone Ltd, Edinburgh, UK P.1457.
- Wilkins, R.J. & Jones, R. 2000. Alternative home-grown protein sources for ruminants in the United Kingdom. Animal Feed Science and Technology 85:23-32.
- Wong, K.H., Li, G.Q., Li, K.M., Razmovski-Naumovski, V. & Chan, K., 2011. Kudzu root: Traditional uses and potential medicinal benefits in diabetes and cardiovascular diseases. Journal of Ethnopharmacology 134: 584–607.

Wu, Y.V. & Nichols, N.N. 2005. Fine grinding and air classification of field pea. Cereal Chemistry 82: 341-344.

- Yadava, R.N. & Singh, A. 1993. A novel flavonone glycoside from the seeds of *Crotalaria laburnifolia*. Fitoterapia 64: 276.
- Yan, Q., Jiang, Z., Yang, S., Deng, W. & Han, L. 2005. A novel homodimeric lectin from *Astragalus mongholicus* with antifungal activity. Archives of Biochemistry and Biophysics 442: 72–81.
- Yang, S.W., Cordell, G.A., Lotter, H., Wagner, H., Mouly, B.C., Rao, A.V.N.A. & Rao, P.S. 1998. Munchiwarin, a prenylated chalcone from *Crotalaria trifoliastrum*. Journal of Natural Products 61: 1274-1276.
- Zhang, Y., Overland, M., Sorensen, M., Penn, M., Mydland, L.T., Shearer, K.D. & Storebakken, T. 2012. Optimal inclusion of lupin and pea protein concentrates in extruded diets for rainbow trout (*Oncorhynchus mykiss*). Aquaculture 344/349: 100-113.